# Concise synthesis of a heptasaccharide antigen found in the cell-wall lipopolysaccharide of *Mycobacterium* gordonae strain 990

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**Abstract** A straight forward synthesis of a heptasaccharide part of the cell-wall lipopolysaccharide of *Mycobacterium gordonae* strain 990, known to have antigenicity, has been achieved in excellent yield. Judicious choice of protecting groups in the intermediates played a significant role throughout the synthesis. Most of the intermediate steps furnished satisfactory yield.

**Keywords** Oligosaccharides · Glycosylations · Antigens · Vaccines · *Mycobacterium gordonae* 990

### Introduction

Since the emergence of acquired immunodeficiency syndrome (AIDS), mycobacterial infection has become a serious concern [1, 2]. In AIDS patients, infections due to non-tubercular mycobacteria (NTM) other than tuberculosis are gradually increasing causing severe problems associated with unnecessary treatments with therapeutics [3, 4]. Among several NTM species isolated from soil and water [5], *Mycobacterium gordonae* (*M. gordonae*) is one of the important organisms, which requires extra attention. Other than soil and water *M. gordonae* can be found in the sputum, urine and gastric juice in human [6]. Earlier it was considered as a friendly organism and often called as "*Mycobacterium aque* or tap-water bacillus" [7]. After-

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wards, a number of infections in the skin, soft tissues, respiratory tract, liver and immunosuppression associated with this species have been reported [8, 9]. In HIV infected patients this particular species causes serious pulmonary infections, which are indistinguishable from the tuberculosis. As a result, treatment with currently available anti-tubercular drugs e.g. isoniazid, pyrazinamide, ethambutol and cycloserine can not cure the patient as *M. gordonae* is resistant to these drugs [8, 9]. Therefore, urgent attention is essential to identify such atypical *Mycobacteria* from other species of *Mycobacteria* for the proper drug administration and thereby AIDS management.

*M. gordonae* 990 strain is a member of atypical NTM, which causes several pulmonary infections in AIDS patients. The cell-wall of this particular strain contains a large number of glycolipids and some of them are antigenic in nature. Brennan *et al.* have isolated and determined the structure of a unique trehalose linked heptasaccharide moiety having antigenic activity (Fig. 1) [10]. As an antigenic oligosaccharide can produce specific immune response in the host by developing specific antibodies, this particular heptasaccharide moiety could also provide useful serodiagnosis of the mycobacterial infection by developing specific antibodies and thus help to design an antibacterial vaccine candidate against this strain.

Although, glycoconjugate vaccines are highly effective to control several bacterial infections, numbers of chemically synthesized carbohydrate vaccines are limited. In most of the glycoconjugate vaccines, the oligosaccharide parts were isolated from natural sources. However, a number of reports appeared in the literature for the development of the synthetic glycoconjugate vaccines against several bacterial infections [11–21]. As cell-wall oligosaccharides having antigenicity are attractive targets for the development of glycoconjugate vaccine candidates, their limited availability



Fig. 1 Structure of the heptasaccharide antigen found in the cell-wall of *M. gordonae* 990

from the natural source set the challenges to the organic chemists to develop concise synthetic protocols for their large scale preparation. In order to develop a specific immune response in the host with the synthetic glycoconjugate vaccine candidate, it is essential to link the synthetic oligosaccharide moiety with a protein through a spacer linkage. Therefore, it is recommended that the reducing end of the synthetic oligosaccharide moiety should contain a temporary protecting group, which can be removed during the process of conjugation with the protein. Although, synthesis of a tetrasaccharide methyl glycoside related to the heptasaccharide moiety was reported earlier [22], total synthesis of the full heptasaccharide with temporary anomeric protecting group was not attempted. In this communication, we present a concise chemical synthesis of the full heptasaccharide antigen found in the cell-wall lipopolysaccharide of M. gordonae strain 990 as its 4methoxyphenyl glycoside (Fig. 2).

### **Results and discussion**

The synthesis of the target heptasaccharide **1** was achieved by judicious functional group manipulations and stereoselective glycosylations. Several differentially protected monosaccharide derivatives (Fig. 3), prepared from commercially available sugars using reported methodologies [23] have been used to construct the target molecule **1**.

Ethyl 4-*O*-benzyl-2-*O*-(4-methoxybenzyl)-3-*O*-methyl-1-thio- $\alpha$ -L-rhamnopyranoside (6) was prepared from ethyl 4-*O*-benzyl-1-thio- $\alpha$ -L-rhamnopyranoside (8) [24] in excellent yield following a tin mediated selective methylation followed by 4-methoxybenzylation under alkaline conditions (Scheme 1).



Fig. 2 Chemical structure of the synthesized heptasaccharide as its 4methoxyphenyl glycoside (1)



Fig. 3 Suitably protected monosaccharide building blocks used for the construction of heptasaccharide (1) as its 4-methoxyphenyl glycoside

Sequential stereoselective glycosylations and protecting group manipulations for the synthesis of target heptasaccharide 1 is presented in Scheme 1 and 2. Compounds 2 and 3 were coupled together stereoselectively to furnish disaccharide derivative 9, which was transformed into the disaccharide acceptor 10 following an earlier reported reaction conditions [23]. N-Iodosuccinimide (NIS)trimethylsilyl trifluoromethanesulfonate (TMSOTf) [25, 26] mediated  $\beta$ -selective glycosylation of disaccharide derivative 10 with thioglycoside 3 afforded trisaccharide derivative 11 in 80% yield, which was benzylated to furnish trisaccharide derivative 12 under a one-pot deacetylationbenzylation protocol reported earlier [27]. Exclusive formation of  $\beta$ -glycosidic linkage in the compound 11 was achieved, due to the presence of an O-acetyl group at the C-2 position of the thioglycoside donor 3, which was confirmed by the signature peaks that appeared in its NMR spectra e. g. δ [5.49 (s, PhCH), 5.27 (br s, H-1<sub>A</sub>), 5.17 (s, PhCH), 4.88 (d, J=8.0 Hz, H-1<sub>B</sub>), 4.91 (d, J=8.0 Hz, H-1<sub>C</sub>) in <sup>1</sup> H NMR and  $\delta$  [103.6 (C-1<sub>C</sub>), 101.6 (Ph*C*H), 100.8 (PhCH), 100.6 (C-1<sub>B</sub>), 97.3 (C-1<sub>A</sub>)] in  $^{13}$ C NMR spectra. Oxidative removal of 4-methoxybenzyl group in compound 12 using DDQ [28] afforded the trisaccharide acceptor 13, which was coupled with 2-O-acetylated thioglycoside donor 5 to furnish tetrasaccharide derivative 14 in 77% yield exploiting neighboring group participation. Signals in the NMR spectra of compound 14 confirmed the  $\alpha$ -selective 1,2-trans glycosylation [ $\delta$  5.30 (br s, 1 H, H-1<sub>A</sub>), 4.97 (br s, 1 H, H-1<sub>D</sub>) in <sup>1</sup>H NMR and  $\delta$  103.4 (C-1<sub>C</sub>), 102.1 (C-1<sub>B</sub>), 101.5 (PhCH), 101.2 (PhCH), 97.8 (C-1<sub>D</sub>), 97.3 (C-1<sub>A</sub>) in <sup>13</sup>C NMR spectra]. Saponification of compound 14 gave tetrasaccharide acceptor 15, which on NIS-TMSOTf mediated glycosylation with the D-xylose



Scheme 1 Reagents: a (1) Bu<sub>2</sub>SnO, toluene, 110°C, 4 h, then iodomethane, TBAB, r t, 48 h; (2) 4-methoxybenzyl chloride, NaOH, DMF, r t, 3 h, 75% in two steps



Scheme 2 Reagents: *a N*-lodosuccinimide, TMSOTf,  $CH_2CI_2$ , MS-4 Å, -40°C, 80% for 11, 77% for 14, 78% for 16; *b* benzyl bromide, NaOH, TBAB, DMF, r t, 3 h, 85%; *c* DDQ,  $CH_2CI_2$ -H<sub>2</sub>O, r t, 2 h, 77% for 13 and 80% for 17; *d* CH<sub>3</sub>ONa, CH<sub>3</sub>OH, r t, 2 h, quantitative

derived thioglycoside donor 7 furnished pentasaccharide derivative **16** in 78% yield. As mentioned earlier stereoselective 1,2-*trans* glycosylation was achieved, due to the presence of an *O*-acetyl group at the C-2 position of compound 7, which was confirmed from the 1D and 2D NMR spectra of compound 16 [ $\delta$  4.10 (d, J=7.2 Hz, H-1<sub>E</sub>,  $\beta$ -D-Xylp) in <sup>1</sup>H NMR and 103.4 (C-1<sub>C</sub>), 102.5 (C-1<sub>B</sub>), 101.9 (C-1<sub>E</sub>), 101.5 (PhCH), 101.3 (PhCH), 99.2 (C-1<sub>D</sub>), 97.2 (C-1<sub>A</sub>) in <sup>13</sup>C NMR]. Compound 16 was allowed to react with DDQ to furnish pentasaccharide acceptor 17 via oxidative removal of 4-methoxybenzyl group (Scheme 2). Compound 17 was allowed to couple with thioglycoside donor 6 under NIS-TMSOTf mediated glycosylation conditions to afford hexasaccharide 18. Although in this case, due to the presence of the 4-methoxybenzyl group, a non-participating functional group at the C-2 position of Lrhamnosyl donor, there was a chance of formation of some β-L-rhamnose linked hexasaccharide derivative together with required compound 18, spectral data of compound 18 confirmed the formation of only the  $\alpha$ -linked required product [signals at  $\delta$  5.51 (s, PhCH), 5.30 (d, J=1.7 Hz, H-1<sub>A</sub>), 5.28 (s, PhC*H*), 5.08 (br s, H-1<sub>D</sub>), 5.05 (br s, H-1<sub>F</sub>), 4.97 (d, J=7.8 Hz, H-1<sub>B</sub>), 4.90 (d, J=8.1 Hz, H-1<sub>C</sub>), 4.70 (d, J=7.2 Hz, H-1<sub>E</sub>) in <sup>1</sup>H NMR, signals at  $\delta$  103.3 (C-1<sub>C</sub>), 102.0 (C-1<sub>B</sub>), 101.2 (2 C, 2 PhCH), 100.7 (C-1<sub>E</sub>), 99.4 (C-1<sub>F</sub>), 98.7 (C-1<sub>D</sub>), 97.1 (C-1<sub>A</sub>) in  $^{13}$ C NMR and proton coupled <sup>13</sup>C NMR spectra [( $J_{C-1/H-1}$  160 Hz, 160 Hz (2  $\beta$ -D-Glcp), 164 Hz (β-D-Xylp), 172 Hz, 172 Hz and 170 Hz (3  $\alpha$ -L-Rhap)]. The values of the coupling constants ( $J_{C-1/H-1}$ ) being above 165 Hz for L-rhamnose moieties confirmed their  $\alpha$ -linkages [22, 29]. Compound 18 was treated with DDQ under similar reaction conditions mentioned earlier to generate hexasaccharide acceptor 19 in 80% yield. Finally, iodonium ion catalyzed  $\alpha$ -selective glycosylation of compound 19 with per-O-acetylated thioglycoside donor 4 furnished heptasaccharide derivative 20 in 85% yield, which on global deprotection of functional groups involving saponification and hydrogenolysis [30] resulted the target heptasaccharide 1 as its 4-methoxyphenyl glycoside. Formation of the required glycosyl linkage in compound 20 was confirmed from its spectral data [signals at  $\delta$  102.2 (C-1<sub>C</sub>), 101.1 (C-1<sub>B</sub>), 100.2 (2 C, PhCH), 99.6 (C-1<sub>F</sub>), 99.4  $(C-1_E)$ , 97.7 (2 C, C-1<sub>D</sub> and C-1<sub>G</sub>), 96.1 (C-1<sub>A</sub>) in <sup>13</sup>C NMR spectrum]. Compound 1 was characterized by its 1D and 2D spectral analysis [signals at  $\delta$  5.39 (br s, 1 H, H-1<sub>G</sub>), 5.33 (br s, 1 H, H-1<sub>A</sub>), 5.26 (br s, 1 H, H-1<sub>D</sub>), 4.92 (br s, 1 H, H-1<sub>F</sub>), 4.70 (d, J=7.5 Hz, 1 H, H-1<sub>B</sub>), 4.63 (d, J=7.5 Hz, 1 H, H-1<sub>C</sub>), 4.54 (d, J=7.2 Hz, 1 H, H-1<sub>E</sub>) in the <sup>1</sup>H NMR and at  $\delta$  105.0 (C-1<sub>E</sub>), 103.9 (C-1<sub>B</sub>), 103.5 (C-1<sub>C</sub>), 101.7 (C-1<sub>F</sub>), 100.7 (C-1<sub>D</sub>), 100.0 (C-1<sub>G</sub>), 99.0 (C-1<sub>A</sub>) in the <sup>13</sup>C NMR spectra] (Scheme 3). Use of functional groups with participating ability (O-acetyl) at C-2 positions of the glycosyl donors resulted in the required stereooutcome in all 1,2-trans glycosylation products. General glycosylation conditions and a similar kind of protecting group strategy were adopted to make this synthetic strategy useful for a scale-up preparation.



Scheme 3 Reagents: *a N*-Iodosuccinimide, TMSOTf,  $CH_2CI_2$ , MS-4 Å, -30°C, 30 min, 74% for 18, 85% for 20; *b* DDQ,  $CH_2CI_2$ -H<sub>2</sub>O, r t, 2 h, 80%; *c* CH<sub>3</sub>ONa, CH<sub>3</sub>OH, r t, 2 h; *d* H<sub>2</sub>, 20% Pd(OH)<sub>2</sub>-C, CH<sub>3</sub>OH, r t, 24 h, 76% in two steps

# Conclusion

In summary, a concise chemical synthesis of the heptasaccharide motif as its 4-methoxyphenyl glycoside found in the cell-wall of *Mycobacterium gordonae* strain 990 has been achieved using general glycosylation conditions and a similar type of protecting group manipulations. This synthetic sequence can also be applied to a scale-up preparation on demand. All glycosylation steps are reasonably fast, stereoselective, high yielding and highly reproducible. Most of the intermediates were solid and characterized with the help of NMR and mass spectral analysis. 4-Methoxy phenyl group can serve as a temporary anomeric protecting group, which can be oxidatively removed using ammonium ceric nitrate (CAN) to couple the heptasaccharide moiety to a carrier protein for the preparation of glycoconjugate antigens.

## Experimental section

*General methods* All the reactions were monitored by thin layer chromatography over silica gel coated TLC plates. The spots on TLC were visualized by warming ceric sulphate  $(2\% \text{ Ce}(SO_4)_2 \text{ in } 2N \text{ H}_2SO_4)$  sprayed plates on a hot plate. Silica gel 230–400 mesh was used for column chromatography. <sup>1</sup>H and <sup>13</sup>C NMR (<sup>1</sup>H coupled and

decoupled), 2D COSY, and HSQC spectra were recorded on Brucker Advance DPX 300 MHz using CDCl<sub>3</sub> and D<sub>2</sub>O as solvents and TMS as internal reference unless stated otherwise. Chemical shift values are expressed in  $\delta$  ppm. ESI-MS were recorded on a MICROMASS QUTTRO II triple quadrupole mass spectrometer. Elementary analysis was carried out on Carlo ERBA-1108 analyzer. Optical rotations were measured at 25°C on a Rudolf Autopol III polarimeter. Commercially available grades of organic solvents of adequate purity are used in many reactions.

Ethvl 4-O-benzvl-2-O-(4-methoxybenzvl)-3-O-methyl-1thio- $\alpha$ -L-rhamnopyranoside (6) To a solution of compound 8 (7.5 g, 25.1 mmol) in toluene (200 mL) was added dibutyltin oxide (7.5 g, 30.1 mmol) and the reaction mixture was allowed to stir at 110°C with azeotropic removal of water for 4 h. The solvents were reduced to half of the volume and iodomethane (6.3 mL, 101.2 mmol) and tetrabutylammonium bromide (1 g) were added to it and the reaction mixture was stirred at room temperature for 48 h. The solvents were removed under reduced pressure and crude product was diluted with CH2Cl2 (150 mL). The organic layer was washed with 1 N aq. HCl, satd. NaHCO<sub>3</sub> and water in succession, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure. To a solution of the crude product in DMF (50 mL) were added powdered NaOH (3 g, 75 mmol), 4-methoxybenzyl chloride (6 mL, 44.2 mmol) and the reaction mixture was allowed to stir at room temperature for 3 h. The reaction mixture was diluted with water (150 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (150 mL). The organic layer was washed with water, dried (Na<sub>2</sub>SO<sub>4</sub>) and evaporated to dryness. The crude product was purified over  $SiO_2$  using hexane-EtOAc (5:1) as eluant to furnish pure compound **6** (8.2 g, 75%); colorless oil;  $[\alpha]_D^{25}$  -47.1 (c 1.5, CHCl<sub>3</sub>); IR (neat): 2925, 2361, 1612, 1512, 1457, 1219, 1099, 1032, 758 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.35–7.26 (m, 7 H, Ar-H), 6.90 (d, J=8.6 Hz, 2 H, Ar-H), 5.24 (br s, 1 H, H-1), 4.94 (d, J=11.2 Hz, 1 H, PhCH<sub>2</sub>), 4.71–4.60 (m, 3 H, PhCH<sub>2</sub>), 3.99 (t, J=7.0 Hz, 1 H, H-4), 3.86-3.85 (m, 1 H, H-5), 3.83 (s, 3 H, OCH<sub>3</sub>), 3.56-3.47 (m, 2 H, H-2 and H-3), 3.38 (s, 3 H, OCH<sub>3</sub>), 2.66-2.52 (m, 2 H, SCH<sub>2</sub>CH<sub>3</sub>), 1.33 (d, J=6.2 Hz, 3 H, CCH<sub>3</sub>), 1.27 (t, J=7.4 Hz, 3 H, SCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 159.3–113.8 (Ar-C), 82.3 (C-1), 81.9 (C-4), 80.6 (C-2), 75.4, 75.1 (2 PhCH<sub>2</sub>), 71.8 (C-3), 68.2 (C-5), 57.4 (OCH<sub>3</sub>), 55.1 (OCH<sub>3</sub>), 25.4 (SCH<sub>2</sub>CH<sub>3</sub>), 17.9 (CCH<sub>3</sub>), 15.1 (SCH<sub>2</sub>CH<sub>3</sub>); ESI-MS: m/z=455.2 [M+Na]<sup>+</sup>; Anal. Calcd. for C<sub>24</sub>H<sub>32</sub>O<sub>5</sub>S (432.20): C, 66.64; H, 7.46; found: C, 66.43; H, 7.70.

4-Methoxyphenyl [2-O-acetyl-4,6-O-benzylidene-3-O-(4-methoxybenzyl)- $\beta$ -D-glucopyranosyl]-(1 $\rightarrow$ 3)-(2-Obenzyl-4,6-O-benzylidine- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-2, 4-di-O-benzyl- $\alpha$ -L-rhamnopyranoside (11) To a solution of compound 10 (4 g, 5 mmol) and compound 3 (2.9 g, 6.1 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (30 mL) was added MS 4 Å (3 g) and the reaction mixture was allowed to stir at room temperature for 30 min under argon. After cooling the reaction mixture to -40°C, N-iodosuccinimide (1.7 g, 7.5 mmol) and TMSOTf (50 µL, 0.27 mmol) were added to it. The reaction mixture was stirred at -40°C for 30 min and quenched with Et<sub>3</sub>N (0.2 mL). The reaction mixture was filtered and washed with CH<sub>2</sub>Cl<sub>2</sub> (30 mL). The organic layer was washed successively with aq. Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> and water, dried ( $Na_2SO_4$ ) and concentrated under reduced pressure to give the crude product, which was purified over SiO<sub>2</sub> using hexane-EtOAc (6:1) as eluant to furnish pure 11 (4.8 g, 80%); colorless solid; m.p. 86–87°C;  $[\alpha]_D^{25}$  –42.7 (c 1.5, CHCl<sub>3</sub>); IR (KBr): 2924, 1749, 1608, 1504, 1407, 1232, 1091, 1030, 754 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.48–7.07 (m, 27 H, Ar-H), 6.88 (d, J=9.0 Hz, 2 H, Ar-H), 6.79-6.74 (m, 4 H, Ar-H), 5.49 (s, 1 H, PhCH), 5.27 (br s, 1 H, H-1<sub>A</sub>), 5.17 (s, 1 H, PhCH), 4.99 (t, J=8.0 Hz, 1 H, H-2<sub>C</sub>), 4.93 (d, J=11.6 Hz, 1 H, PhCH<sub>2</sub>), 4.91 (d, J=8.0 Hz, 1 H, H-1<sub>C</sub>), 4.88 (d, J=8.0 Hz, 1 H, H-1<sub>B</sub>), 4.85 (d, J= 12.1 Hz, 1 H, PhCH<sub>2</sub>), 4.75 (d, J=12.1 Hz, 1 H, PhCH<sub>2</sub>), 4.73–4.68 (m, 3 H, PhC $H_2$ ), 4.59 (d, J= 11.8 Hz, 1 H, PhCH<sub>2</sub>), 4.36–4.28 (m, 3 H, H-3<sub>A</sub>, H-6<sub>aC</sub> and PhCH<sub>2</sub>), 4.14– 4.09 (m, 1 H, H-6<sub>aB</sub>), 3.96–3.95 (m, 1 H, H-2<sub>A</sub>), 3.90–3.77 (m, 3 H, H-3<sub>B</sub>, H-3<sub>C</sub> and H-6<sub>bC</sub>), 3.76, 3.74 (2 s, 6 H, 2 OCH<sub>3</sub>), 3.71–3.51 (m, 5 H, H-4<sub>A</sub>, H-4<sub>B</sub>, H-5<sub>B</sub>, H-5<sub>C</sub> and H-6<sub>bB</sub>), 3.43 (t, J=8.1 Hz, 1 H, H-2<sub>B</sub>), 3.36–3.28 (m, 2 H, H-4<sub>C</sub> and H-5<sub>A</sub>), 1.78 (s, 3 H, COCH<sub>3</sub>), 1.27 (d, J=6.3 Hz, 3 H, CCH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 169.0 (COCH<sub>3</sub>), 159.2–113.6 (Ar-C), 103.6 (C-1<sub>C</sub>), 101.6 (PhCH), 100.8 (PhCH), 100.6 (C-1<sub>B</sub>), 97.3 (C-1<sub>A</sub>), 82.7 (C-2<sub>B</sub>), 81.5 (C-5<sub>C</sub>), 80.7 (C-5<sub>B</sub>), 79.7 (C-3<sub>C</sub>), 78.9 (C-4<sub>B</sub>), 78.4 (C-3<sub>B</sub>), 78.1 (C-2<sub>A</sub>), 77.0 (C-2<sub>C</sub>), 75.4 (PhCH<sub>2</sub>), 74.5 (PhCH<sub>2</sub>), 73.7 (PhCH<sub>2</sub>), 73.4 (C-3<sub>A</sub>), 72.9 (PhCH<sub>2</sub>), 68.7 (C-6<sub>B</sub>), 68.6 (C-6<sub>C</sub>), 68.5 (C-4<sub>A</sub>), 66.3 (C-5<sub>A</sub>), 65.9 (C-4<sub>C</sub>), 55.5 (OCH<sub>3</sub>), 55.1 (OCH<sub>3</sub>), 20.7 (COCH<sub>3</sub>), 18.0 (CCH<sub>3</sub>); ESI-MS: m/z= 1225.6 [M+Na]<sup>+</sup>; Anal. Calcd. for C<sub>70</sub>H<sub>74</sub>O<sub>18</sub> (1202.48): C, 69.87; H, 6.20; found: C, 69.70; H, 6.46.

4-Methoxyphenyl [2-O-benzyl-4,6-O-benzylidene-3-O-(4-methoxybenzyl)- $\beta$ -D-glucopyranosyl]-(1 $\rightarrow$ 3)-(2-O-benzyl-4,6-O-benzylidine- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-2,4-di-Obenzyl- $\alpha$ -L-rhamnopyranoside (12) To a solution of compound 11 (4.5 g, 3.74 mmol) in DMF (50 mL) were added benzyl bromide (0.9 mL, 7.6 mmol), TBAB (0.2 g), powdered NaOH (450 mg, 11.2 mmol) and the reaction mixture was allowed to stir at room temperature for 3 h. The reaction mixture was diluted with water (150 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (100 mL). The organic layer was washed with water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated to give the crude product, which was purified over SiO<sub>2</sub> using hexane-EtOAc (4:1) as eluant to furnish pure 12 (4 g, 85%); colorless solid; m.p. 67–69°C;  $[\alpha]_D^{25}$  –31.3 (c 1.5, CHCl<sub>3</sub>); IR (KBr): 2926, 1720, 1613, 1508, 1456, 1367, 1247, 1219, 1080, 827, 739, 699  $\text{cm}^{-1}$ ; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): § 7.51–7.17 (m, 32 H, Ar-H), 6.95 (d, J=9.0 Hz, 2 H, Ar-H), 6.80-6.77 (m, 4 H, Ar-H), 5.55 (s, 1 H, PhCH), 5.33 (d, J=1.7 Hz, 1 H, H-1<sub>A</sub>), 5.31 (s, 1 H, PhCH), 5.07–4.95 (m, 2 H, PhCH<sub>2</sub>), 4.93 (d, J=8.0 Hz, 1 H, H-1<sub>C</sub>), 4.84 (d, J=7.7 Hz, 1 H, H-1<sub>B</sub>), 4.82-4.63 (m, 7 H, PhCH<sub>2</sub>), 4.39–4.34 (m, 3 H, H-3<sub>A</sub>, H-6<sub>aC</sub> and PhCH<sub>2</sub>), 4.27–4.20 (m, 1 H, H-6<sub>aB</sub>), 4.02 (t, J=8.5 H, 1 H,  $H-3_{C}$ ), 3.99–3.97 (m, 1 H,  $H-2_{A}$ ), 3.86–3.78 (m, 2 H,  $H-3_{B}$ and H-4<sub>A</sub>), 3.79 (s, 6 H, 2 OCH<sub>3</sub>), 3.76–3.72 (m, 1 H,  $H-6_{bC}$ ), 3.70–3.63 (m, 4 H,  $H-4_{B}$ ,  $H-4_{C}$ ,  $H-5_{B}$  and  $H-6_{bB}$ ), 3.54 (t, J=8.0 Hz, H-2<sub>C</sub>), 3.51 (t, J=7.6 Hz, 1 H, H-2<sub>B</sub>), 3.48–3.40 (m, 1 H, H-5<sub>A</sub>), 3.38–3.30 (m, 1 H, H-5<sub>C</sub>), 1.28 (d, J=6.0 Hz, 3 H, CCH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 159.4–113.6 (Ar-C), 103.5 (C-1<sub>C</sub>), 102.2 (PhCH), 101.5 (PhCH), 100.8 (C-1<sub>B</sub>), 97.4 (C-1<sub>A</sub>), 83.3 (C-2<sub>B</sub>), 82.2 (C-5<sub>C</sub>), 81.5 (C-5<sub>B</sub>), 81.2 (C-3<sub>A</sub>), 80.2 (C-3<sub>C</sub>), 79.7 (C-4<sub>B</sub>), 78.5 (C-3<sub>B</sub>), 78.2 (C-2<sub>A</sub>), 77.0 (C-2<sub>C</sub>), 75.1 (PhCH<sub>2</sub>), 74.5 (2 C, PhCH<sub>2</sub>), 74.0 (PhCH<sub>2</sub>), 73.7 (PhCH<sub>2</sub>), 68.9 (2 C, C-6<sub>B</sub> and C-6<sub>C</sub>), 68.6 (C-4<sub>A</sub>), 66.2 (C-5<sub>A</sub>), 65.7 (C-4<sub>C</sub>), 55.5 (OCH<sub>3</sub>), 55.0 (OCH<sub>3</sub>), 17.9 (CCH<sub>3</sub>); ESI-MS: m/z=1268.5 [M  $+NH_4$ ]<sup>+</sup>; Anal. Calcd. for C<sub>75</sub>H<sub>78</sub>O<sub>17</sub> (1250.52): C, 71.98; H, 6.28; found: C, 71.78; H, 6.54.

4-Methoxyphenyl (2-O-benzyl-4, 6-O-benzylidene- $\beta$ -Dglucopyranosyl)- $(1 \rightarrow 3)$ -(2-O-benzyl-4, 6-O-benzylidine- $\beta$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -2,4-di-O-benzyl- $\alpha$ -L-rhamnopyranoside (13) To a solution of compound 12 (3.8 g, 3 mmol) in CH<sub>2</sub>Cl<sub>2</sub> and water (50 mL, 1:1), was added DDQ (820 mg, 3.6 mmol) and the reaction mixture was allowed to stir at room temperature for 2 h. The reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (50 mL) and the organic layer was washed successively with satd. aq NaHCO3 and water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure to give the crude product, which was purified over SiO<sub>2</sub> using hexane-EtOAc (4:1) to furnish pure 13 (2.6 g, 77%); colorless solid; m.p. 96–98°C; [α]<sub>D</sub><sup>25</sup> –40.3 (*c* 1.5, CHCl<sub>3</sub>); IR (KBr): 2874, 1632, 1506, 1457, 1378, 1220, 1091, 1033, 998, 745, 698 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.50-7.07 (m, 30 H, Ar-H), 6.92 (d, J=9.0 Hz, 2 H, Ar-H), 6.78 (d, J=9.0 Hz, 2 H, Ar-H), 5.52 (s, 1 H, PhCH), 5.32 (s, 1 H, PhCH), 5.30 (d, J=1.6 Hz, 1 H, H-1<sub>A</sub>), 5.01  $(d, J=8.0 \text{ Hz}, 1 \text{ H}, \text{H}-1_{\text{C}}), 4.99 (d, J=7.9 \text{ Hz}, 1 \text{ H}, \text{H}-1_{\text{B}}),$ 4.96 (d, J=11.7 Hz, 1 H, PhCH<sub>2</sub>), 4.89 (d, J=11.7 Hz, 1 H, PhCH<sub>2</sub>), 4.85 (d, J=12.0 Hz, 1 H, PhCH<sub>2</sub>), 4.78 (d, J= 11.7 Hz, 1 H, PhCH<sub>2</sub>), 4.71–4.65 (m, 3 H, PhCH<sub>2</sub>), 4.37– 4.32 (m, 3 H, H-3<sub>A</sub>, H-6<sub>aC</sub> and PhCH<sub>2</sub>), 4.24–4.19 (m, 1 H, H-6<sub>aB</sub>), 4.01 (t, J=8.9 Hz, 1 H, H-3<sub>C</sub>), 3.97–3.95 (m, 1 H, H-2<sub>A</sub>), 3.85–3.76 (m, 1 H, H-4<sub>A</sub>), 3.75 (s, 3 H, OCH<sub>3</sub>), 3.73–3.69 (m, 1 H, H-3<sub>B</sub>), 3.67–3.57 (m, 3 H, H-4<sub>C</sub>, H-6<sub>bB</sub>

and H-6<sub>b</sub>C), 3.55–3.48 (m, 3 H, H-2<sub>C</sub>, H-4<sub>B</sub> and H-5<sub>B</sub>), 3.47–3.28 (m, 3 H, H-2<sub>B</sub>, H-5<sub>A</sub> and H-5<sub>C</sub>), 1.27 (d, J= 6.0 Hz, 3 H, CCH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  150.2– 114.5 (Ar-C), 103.5 (C-1<sub>C</sub>), 102.4 (C-1<sub>B</sub>), 101.5 (PhCH), 101.4 (PhCH), 97.2 (C-1<sub>A</sub>), 83.1 (C-5<sub>B</sub>), 82.2 (C-2<sub>B</sub>), 81.5 (C-5<sub>C</sub>), 80.5 (C-4<sub>B</sub>), 79.5 (C-3<sub>A</sub>), 78.5 (C-2<sub>A</sub>), 78.4 (C-3<sub>C</sub>), 76.8 (C-2<sub>C</sub>), 75.1 (PhCH<sub>2</sub>), 74.4 (2 C, 2 PhCH<sub>2</sub>), 73.6 (PhCH<sub>2</sub>), 73.3 (C-3<sub>B</sub>), 68.7 (2 C, C-6<sub>B</sub> and C-6<sub>C</sub>), 68.5 (C-4<sub>A</sub>), 66.2 (C-5<sub>A</sub>), 65.7 (C-4<sub>C</sub>), 55.4 (OCH<sub>3</sub>), 17.9 (CCH<sub>3</sub>); ESI-MS: *m*/*z*=1153.5 [M+Na]<sup>+</sup>; Anal. Calcd. for C<sub>67</sub>H<sub>70</sub>O<sub>16</sub> (1130.46): C, 71.13; H, 6.24; found: C, 70.92; H, 6.50.

4-Methoxyphenyl [2-O-acetyl-4-O-benzyl-3-O-(4-methoxybenzyl)- $\alpha$ -L-rhamnopyranosyl]- $(1 \rightarrow 3)$ -(2-O-benzyl-4, 6-O $benzylidene-\beta$ -D-glucopyranosyl)- $(1\rightarrow 3)$ -(2-O-benzyl-4,6-O-benzylidine- $\beta$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -2,4-di-Obenzyl- $\alpha$ -L-rhamnopyranoside (14) To a solution of compound 13 (2.5 g, 2.2 mmol) and compound 5 (1.2 g, 2.6 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (25 mL) was added MS 4 Å (2 g) and the reaction mixture was stirred at room temperature for 30 min under argon. The reaction mixture was cooled to -40°C and N-iodosuccinimide (700 mg, 3.1 mmol) and TMSOTf (20 µL, 0.12 mmol) were added to it. The mixture was stirred at -40°C for 30 min and quenched with Et<sub>3</sub>N (0.1 mL). The reaction mixture was filtered and washed with CH<sub>2</sub>Cl<sub>2</sub> (50 mL). The filtrate was successively washed with 10% aq Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> and water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure to give crude product, which was purified over SiO<sub>2</sub> using hexane-EtOAc (4:1) as eluant to furnish pure 14 (2.6 g, 77%); colorless solid; m.p. 100–101°C; [α]<sub>D</sub><sup>25</sup> –38.6 (*c* 1.5, CHCl<sub>3</sub>); IR (KBr): 2926, 2361, 1741, 1705, 1510, 1459, 1374, 1238, 1090, 754, 697 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.46–7.09 (m, 37 H, Ar-H), 6.89 (d, J=9.0 Hz, 2 H, Ar-H), 6.78-6.74 (m, 4 H, Ar-H), 5.49 (s, 1 H, PhCH), 5.38-5.36 (m, 1 H, H-2<sub>D</sub>), 5.30 (br s, 1 H, H-1<sub>A</sub>), 5.25 (s, 1 H, PhCH), 5.02-4.98 (m, 3 H, H-1<sub>B</sub>, H-1<sub>C</sub> and PhCH<sub>2</sub>), 4.97 (br s, 1 H, H-1<sub>D</sub>), 4.94–4.77 (m, 5 H, PhCH<sub>2</sub>), 4.70–4.58 (m, 4 H, PhCH<sub>2</sub>), 4.51 (d, J=11.2 Hz, 1 H, PhCH<sub>2</sub>), 4.38-4.31 (m, 4 H, H-3<sub>A</sub>, H-4<sub>D</sub>, H-6<sub>aC</sub> and PhCH<sub>2</sub>), 4.24–4.17 (m, 1 H,  $H-6_{aB}$ ), 4.08–4.0 (m, 2 H,  $H-3_{C}$  and  $H-4_{C}$ ), 3.97–3.95 (m, 1 H, H-2<sub>A</sub>), 3.85–3.79 (m, 3 H, H-3<sub>B</sub>, H-3<sub>D</sub> and H-6<sub>bC</sub>), 3.74, 3.72 (2 s, 6 H, 2 OCH<sub>3</sub>), 3.70–3.59 (m, 4 H, H-4<sub>A</sub>, H- $5_{B}$ , H- $5_{D}$  and H- $6_{bB}$ ), 3.58–3.40 (m, 3 H, H- $2_{B}$ , H- $2_{C}$  and H-4<sub>B</sub>), 3.35–3.20 (m, 2 H, H-5<sub>A</sub> and H-5<sub>C</sub>), 2.03 (s, 3 H, COCH<sub>3</sub>), 1.27 (d, J=6.0 Hz, 3 H, CCH<sub>3</sub>), 0.93 (d, J= 6.0 Hz, 3 H, CCH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 169.7 (COCH<sub>3</sub>), 159.2–113.7 (Ar-C), 103.4 (C-1<sub>C</sub>), 102.1 (C-1<sub>B</sub>), 101.5 (PhCH), 101.2 (PhCH), 97.8 (C-1<sub>D</sub>), 97.3 (C-1<sub>A</sub>), 83.3 (C-2<sub>B</sub>), 82.6 (C-2<sub>C</sub>), 81.5 (C-5<sub>B</sub>), 80.1 (C-3<sub>C</sub>), 79.6  $(C-5_C)$ , 79.0  $(C-5_D)$ , 78.5  $(C-2_A)$ , 77.8 (2 C, C-3<sub>B</sub> and C-3<sub>D</sub>), 76.9 (C-3<sub>A</sub>), 76.3 (C-4<sub>B</sub>), 75.1 (PhCH<sub>2</sub>), 74.9 (PhCH<sub>2</sub>), 74.6 (PhCH<sub>2</sub>), 74.5 (PhCH<sub>2</sub>), 73.7 (PhCH<sub>2</sub>), 71.4 (PhCH<sub>2</sub>), 68.9 (2 C, C-6<sub>B</sub> and C-6<sub>C</sub>), 68.7 (C-2<sub>D</sub>), 68.6 (C-4<sub>A</sub>), 67.5 (C-4<sub>C</sub>), 66.2 (C-5<sub>A</sub>), 66.0 (C-4<sub>D</sub>), 55.4 (OCH<sub>3</sub>), 55.0 (OCH<sub>3</sub>), 20.9 (COCH<sub>3</sub>), 17.9 (CCH<sub>3</sub>), 17.5 (CCH<sub>3</sub>); ESI-MS: m/z=1546.7 [M+NH<sub>4</sub>]<sup>+</sup>; Anal. Calcd. for C<sub>90</sub>H<sub>96</sub>O<sub>22</sub> (1528.64): C, 70.66; H, 6.33; found: C, 70.48; H, 6.55.

4-Methoxyphenyl [4-O-benzyl-3-O-(4-methoxybenzyl)- $\alpha$ -L-rhamnopyranosyl]- $(1 \rightarrow 3)$ -(2-O-benzyl-4, 6-O-benzylidene- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-(2-O-benzyl-4,6-O-benzylidine- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-2,4-di-O-benzyl- $\alpha$ -L-rhamnopyranoside (15) A solution of the compound 14 (2.5 g. 1.6 mmol) in 0.1 M CH<sub>3</sub>ONa in CH<sub>3</sub>OH (50 mL) was allowed to stir at room temperature for 2 h, neutralized with Amberlite IR-120  $(H^+)$  resin, filtered and concentrated. The crude product was passed through a short pad of SiO<sub>2</sub> using hexane-EtOAc (3:1) to give pure compound 15 (2.4 g, quantitative); colorless solid; m.p. 92–94°C;  $[\alpha]_D^{25}$  –55.6 (c 1.5, CHCl<sub>3</sub>); IR (KBr): 2927, 1620, 1508, 1457, 1377, 1220, 1093, 1036, 744, 697 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): § 7.47-7.01 (m, 37 H, Ar-H), 6.93 (d, J=9.0 Hz, Ar-H), 6.81-6.76 (m, 4 H, Ar-H), 5.50 (s, 1 H, PhCH), 5.31 (d, J=1.7 Hz, 1 H, H-1<sub>A</sub>), 5.28 (s, 1 H, PhCH), 5.06 (br s, 1 H, H-1<sub>D</sub>), 4.98 (2 d, J=7.7 Hz, 2 H, H-1<sub>B</sub> and H-1<sub>C</sub>), 4.95-4.75 (m, 5 H, PhCH<sub>2</sub>), 4.70 (d, J=12.0 Hz, 1 H, PhCH<sub>2</sub>), 4.63–4.47 (m, 5 H, PhCH<sub>2</sub>), 4.37–4.31 (m, 3 H, H-3<sub>A</sub>, H-6<sub>aC</sub> and PhCH<sub>2</sub>), 4.24–4.19 (m, 1 H, H-6<sub>aB</sub>), 4.03– 3.94 (m, 3 H, H-2<sub>A</sub>, H-3<sub>C</sub> and H-3<sub>D</sub>), 3.82–3.76 (m, 2 H, H-3<sub>B</sub> and H-4<sub>D</sub>), 3.75, 3.73 (2 s, 6 H, 2 OCH<sub>3</sub>), 3.72–3.70 (m, 1 H, H-6<sub>bC</sub>), 3.69–3.60 (m, 3 H, H-2<sub>D</sub>, H-4<sub>A</sub> and H-6<sub>bB</sub>), 3.58–3.32 (m, 6 H, H-2<sub>B</sub>, H-4<sub>B</sub>, H-4<sub>C</sub>, H-5<sub>A</sub>, H-5<sub>B</sub> and H-5<sub>D</sub>), 3.30-3.22 (m, 2 H, H-2<sub>C</sub> and H-5<sub>C</sub>), 1.27 (d, *J*=6.0 Hz, 3 H, CC*H*<sub>3</sub>), 0.87 (d, *J*=6.0 Hz, 3 H, CC*H*<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 159.2–113.8 (Ar-C), 103.4 (C-1<sub>C</sub>), 102.5 (C-1<sub>B</sub>), 101.3 (2 C, 2 PhCH), 99.5 (C-1<sub>D</sub>), 97.1 (C-1<sub>A</sub>), 83.2 (C-2<sub>B</sub>), 82.5 (C-2<sub>C</sub>), 81.5 (C-5<sub>B</sub>), 80.1 (C-3<sub>C</sub>), 79.5 (C-5<sub>C</sub>), 79.4 (C-4<sub>A</sub>), 79.2 (C-2<sub>A</sub>), 78.4 (C-3<sub>D</sub>), 78.2 (C-3<sub>B</sub>), 76.8 (2 C, C-3<sub>A</sub> and C-4<sub>B</sub>), 75.2, 74.9, 74.6, 74.5, 73.6 (5 PhCH<sub>2</sub>), 68.8 (2 C, C-6<sub>B</sub> and  $C-6_{C}$ ), 68.5 (C-2<sub>D</sub>), 68.4 (C-5<sub>D</sub>), 67.2 (C-4<sub>C</sub>), 66.2 (C-5<sub>A</sub>), 66.0 (C-4<sub>D</sub>), 55.4 (OCH<sub>3</sub>), 55.0 (OCH<sub>3</sub>), 17.9, 17.3 (2 CCH<sub>3</sub>); ESI-MS: m/z=1504.6 [M+NH<sub>4</sub>]<sup>+</sup>; Anal. Calcd. for C<sub>88</sub>H<sub>94</sub>O<sub>21</sub> (1486.63): C, 71.05; H, 6.37; found: C, 70.84; H, 6.62.

4-Methoxyphenyl (2,3,4-tri-O-acetyl- $\beta$ -D-xylopyranosyl)-(1 $\rightarrow$ 2)-[4-O-benzyl-3-O-(4-methoxybenzyl)- $\alpha$ -L-rhamnopyranosyl]-(1 $\rightarrow$ 3)-(2-O-benzyl-4,6-O-benzylidene- $\beta$ -Dglucopyranosyl)-(1 $\rightarrow$ 3)-(2-O-benzyl-4,6-O-benzylidine- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-2,4-di-O-benzyl- $\alpha$ -L-rhamnopyranoside (16) To a solution of compound 15 (2.3 g, 1.5 mmol) and compound 7 (0.6 g, 1.9 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (20 mL) was added MS 4 Å (1 g) and the reaction mixture was stirred at room temperature for 30 min under argon. The reaction mixture was cooled to -40°C and Niodosuccinimide (500 mg, 2.2 mmol) and TMSOTf (15 µL, 0.08 mmol) were added to it. The mixture was stirred at the same temperature for 30 min and guenched with Et<sub>3</sub>N (0.1 mL). The reaction mixture was filtered and washed with CH<sub>2</sub>Cl<sub>2</sub> (50 mL). The organic layer was washed successively with 10% aq Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> and water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure to give crude product, which was purified over SiO<sub>2</sub> using hexane-EtOAc (5:1) as eluant to furnish pure 16 (2 g, 78%); colorless solid; m.p. 86-88°C;  $\left[\alpha\right]_{D}^{25}$  -53.6 (c 1.5, CHCl<sub>3</sub>); IR (KBr): 2927, 2859, 1754, 1509, 1457, 1371, 1247, 1222, 1179, 1092, 1040, 1000, 828, 749, 699 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.47-7.01 (m, 37 H, Ar-H), 6.90 (d, J=9.0 Hz, 2 H, Ar-H), 6.82-6.75 (m, 4 H, Ar-H), 5.50 (s, 1 H, PhCH), 5.32 (S, 1 H, PhCH), 5.30 (d, J=1.6 Hz, 1 H, H-1<sub>A</sub>), 5.10 (br s, 1 H, H-1<sub>D</sub>), 5.02 (2 d, J=7.7 Hz, 2 H, H-1<sub>B</sub> and H-1<sub>C</sub>), 4.97 (d, J=11.7 Hz, 1 H, PhCH<sub>2</sub>), 4.93–4.90 (m, 2 H, H-2<sub>E</sub> and PhCH<sub>2</sub>), 4.88–4.68 (m, 6 H, PhCH<sub>2</sub>), 4.65–4.58 (m, 2 H, H-3<sub>E</sub> and PhCH<sub>2</sub>), 4.55–4.44 (m, 3 H, PhCH<sub>2</sub>), 4.41 (d, J=7.2 Hz, 1 H, H-1<sub>E</sub>), 4.35–4.23 (m, 4 H, H-3<sub>A</sub>, H-3<sub>D</sub>, H-4<sub>E</sub> and H-6<sub>aC</sub>), 3.97 (t, J=8.0 Hz, 1 H, H-3<sub>C</sub>), 3.96–3.88 (m, 2 H, H-2<sub>A</sub> and H-5<sub>aE</sub>), 3.85-3.77 (m, 3 H, H-2<sub>D</sub>, H-4<sub>A</sub> and H-5<sub>bE</sub>), 3.75 (s, 6 H, 2 OCH<sub>3</sub>), 3.74–3.70 (m, 2 H, H-5<sub>A</sub> and H-6<sub>bC</sub>), 3.68–3.58 (m, 3 H, H-3<sub>B</sub>, H-4<sub>D</sub> and H-6<sub>aB</sub>), 3.57– 3.50 (m, 1 H, H-4<sub>B</sub>), 3.49–3.35 (m, 4 H, H-2<sub>B</sub>, H-4<sub>C</sub>, H-5<sub>D</sub> and H-6<sub>bB</sub>), 3.32-22 (m, 2 H, H-2<sub>C</sub> and H-5<sub>C</sub>), 2.82–2.72 (m, 1 H, H-5<sub>B</sub>), 2.03, 2.01 (2 s, 9 H, 3 COCH<sub>3</sub>), 1.25 (d, J=9.0 Hz, 3 H, CCH<sub>3</sub>), 0.85 (d, J=9.0 Hz, 3 H, CCH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 169.9, 169.8 (2 C) (3 COCH<sub>3</sub>), 159.5–113.7 (Ar-C), 103.4 (C-1<sub>C</sub>), 102.5 (C-1<sub>B</sub>), 101.9 (C-1<sub>E</sub>), 101.5 (PhCH), 101.3 (PhCH), 99.2 (C-1<sub>D</sub>), 97.2 (C-1<sub>A</sub>), 83.2 (C-2<sub>B</sub>), 82.5 (C-5<sub>C</sub>), 81.5 (C-5<sub>B</sub>), 80.5 (C-2<sub>C</sub>), 79.4 (2 C, C-4<sub>A</sub> and C-4<sub>B</sub>), 78.5 (C-2<sub>A</sub>), 78.0 (C-4<sub>D</sub>), 76.9 (C-3<sub>A</sub>), 75.0 (2 C, PhCH<sub>2</sub>), 74.4 (PhCH<sub>2</sub>), 74.2 (PhCH<sub>2</sub>), 73.6 (PhCH<sub>2</sub>), 72.2 (PhCH<sub>2</sub>), 71.2 (C-4<sub>E</sub>), 71.0 (C-2<sub>E</sub>), 70.6 (C-3<sub>E</sub>), 69.5 (C-2<sub>D</sub>), 69.3 (C-3<sub>B</sub>), 69.0 (2 C, C-5<sub>E</sub> and C-6<sub>C</sub>), 68.8 (C-5<sub>D</sub>), 68.6 (C-3<sub>D</sub>), 67.8 (C-3<sub>C</sub>), 66.2 (C-5<sub>A</sub>), 66.0 (C-4<sub>C</sub>), 61.3 (C-6<sub>B</sub>), 55.5, 55.0 (2 OCH<sub>3</sub>), 20.6 (3 C, 3 COCH<sub>3</sub>), 17.9, 17.3 (2 CCH<sub>3</sub>); ESI-MS: m/z= 1762.8  $[M+NH_4]^+$ ; Anal. Calcd. for C<sub>99</sub>H<sub>108</sub>O<sub>28</sub> (1744.70): C, 68.11; H, 6.24; found: C, 67.92; H, 6.46.

4-Methoxyphenyl (2,3,4-tri-O-acetyl- $\beta$ -D-xylopyranosyl)-(1 $\rightarrow$ 2)-(4-O-benzyl- $\alpha$ -L-rhamnopyranosyl)-(1 $\rightarrow$ 3)-(2-O-benzyl-4,6-O-benzylidene- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-(2-O-benzyl-4,6-O-benzylidine- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-2,4-di-O-benzyl- $\alpha$ -L-rhamnopyranoside (17) To a solution of compound **16** (1.8 g, 1 mmol) in CH<sub>2</sub>Cl<sub>2</sub> and water (30 mL, 1:1), was added DDQ (270 mg, 1.2 mmol) and the reaction mixture was stirred at room temperature for 2 h. The reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (30 mL) and the organic layer was washed in succession with satd. aq NaHCO<sub>3</sub> and water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure to give the crude product, which was purified over SiO<sub>2</sub> using hexane-EtOAc (3:1) as eluant to furnish pure 17 (1.3 g, 80%); colorless solid; m.p. 104-106°C; [α]<sub>D</sub><sup>25</sup> -36.9 (*c* 1.5, CHCl<sub>3</sub>); IR (KBr): 2925, 1749, 1605, 1375, 1224, 1086, 761 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): § 7.47–7.02 (m, 35 H, Ar-H), 6.88 (d, J=9.0 Hz, 2 H, Ar-H), 6.78 (d, J=9.0 Hz, 2 H, Ar-H), 5.50 (s, 1 H, PhCH), 5.30 (s, 1 H, PhCH), 5.29 (br s, 1 H, H-1<sub>A</sub>), 5.05 (br s, 1 H, H-1<sub>D</sub>), 5.02–4.88 (m, H-1<sub>B</sub>, H-1<sub>C</sub> and PhCH<sub>2</sub>), 4.85-4.72 (m, 3 H, H-2<sub>E</sub> and PhCH<sub>2</sub>), 4.70-4.66 (m, 2 H, PhCH<sub>2</sub>), 4.64–4.50 (m, 4 H, H-3<sub>E</sub> and PhCH<sub>2</sub>), 4.35–4.27  $(m, 3 H, H-3_A, H-3_D \text{ and } H-4_E), 4.25-4.17 (m, 1 H, H-6_{aC}),$ 4.10 (d, J=7.2 Hz, 1 H, H-1<sub>E</sub>), 4.00 (t, J=7.9 Hz, 1 H, H-3<sub>C</sub>), 3.94–3.92 (m, 1 H, H-2<sub>A</sub>), 3.87–3.76 (m, 5 H, H-3<sub>B</sub>, H-4<sub>A</sub>, H-4<sub>C</sub>, H-5<sub>D</sub> and H-5<sub>aE</sub>), 3.74 (s, 3 H, OCH<sub>3</sub>), 3.70-3.60 (m, 4 H, H-2<sub>D</sub>, H-5<sub>B</sub>, H-5<sub>bE</sub> and H-6<sub>bC</sub>), 3.58–3.50 (m, 2 H, H-4<sub>B</sub> and H-6<sub>aB</sub>), 3.48-3.35 (m, 3 H, H-2<sub>B</sub>, H-2<sub>C</sub> and H-5<sub>A</sub>), 3.32–3.23 (m, 1 H, H-5<sub>C</sub>), 3.07 (t, J=7.9 Hz, 1 H, H-4<sub>D</sub>), 2.68–2.58 (m, 1 H, H-6<sub>bB</sub>), 2.06, 2.05, 2.03 (3 s, 9 H, 3 COCH<sub>3</sub>), 1.24 (d, J=6.0 Hz, 3 H, CCH<sub>3</sub>), 0.80 (d, J=6.0 Hz, 3 H, CCH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 169.6, 169.5, 169.2 (3 COCH<sub>3</sub>), 154.8–114.4 (Ar-C), 103.2 (C-1<sub>C</sub>), 102.4 (C-1<sub>B</sub>), 102.1 (C-1<sub>E</sub>), 101.5 (Ph*C*H), 101.2 (PhCH), 98.7 (C-1<sub>D</sub>), 97.1 (C-1<sub>A</sub>), 83.0 (C-2<sub>B</sub>), 82.8 (C-5<sub>C</sub>), 81.9 (C-4<sub>D</sub>), 81.3 (C-2<sub>C</sub>), 80.3 (C-5<sub>B</sub>), 79.3 (C-4<sub>A</sub>), 79.2 (C-4<sub>B</sub>), 78.4 (C-2<sub>A</sub>), 78.1 (C-2<sub>D</sub>), 77.2 (C-3<sub>A</sub>), 74.9 (PhCH<sub>2</sub>), 74.4 (2 C, PhCH<sub>2</sub>), 74.3 (PhCH<sub>2</sub>), 73.5 (PhCH<sub>2</sub>), 71.3 (C-4<sub>E</sub>), 71.2 (C-3<sub>C</sub>), 70.9 (C-2<sub>E</sub>), 70.7 (C-3<sub>E</sub>), 69.3 (C-3<sub>B</sub>), 69.2 (C-5<sub>D</sub>), 68.9 (2 C, C-5<sub>E</sub> and C-6<sub>C</sub>), 68.4  $(C-3_D)$ , 66.9  $(C-5_A)$ , 66.0  $(C-4_C)$ , 61.4  $(C-6_B)$ , 55.3 (OCH<sub>3</sub>), 20.5 (3 C, 3 COCH<sub>3</sub>), 17.8, 17.2 (2 CCH<sub>3</sub>); ESI-MS: m/z=1642.7 [M+NH<sub>4</sub>]<sup>+</sup>; Anal. Calcd. for C<sub>91</sub>H<sub>100</sub>O<sub>27</sub> (1624.64): C, 67.23; H, 6.20; found: C, 67.02; H, 6.44.

4-Methoxyphenyl [4-O-benzyl-3-O-methyl-2-O-(4-methoxybenzyl)- $\alpha$ -L-rhamnopyranosyl]- $(1 \rightarrow 3)$ -[(2,3,4-tri-O-acetyl- $\beta$ -D-xvlopyranosyl)-(1 $\rightarrow$ 2)]-(4-O-benzyl- $\alpha$ -L-rhamnopyranosyl)- $(1 \rightarrow 3)$ -(2-O-benzyl-4, 6-O-benzylidene- $\beta$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -(2-O-benzyl-4, 6-O-benzylidine- $\beta$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -2,4-di-O-benzyl- $\alpha$ -L-rhamnopyranoside (18) To a solution of compound 17 (1.2 g, 0.74 mmol) and compound 6 (390 mg, 0.9 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (20 mL) was added MS 4 Å (1 g) and the reaction mixture was stirred at room temperature for 30 min under argon. The reaction mixture was cooled to -30°C and N-iodosuccinimide (225 mg, 1 mmol) and TMSOTf (10 µL, 0.05 mmol) were added to it. The mixture was stirred at the same temperature for 30 min and quenched with Et<sub>3</sub>N (0.1 mL). The reaction mixture was filtered and washed with CH<sub>2</sub>Cl<sub>2</sub> (50 mL). The organic layer was washed successively with 10% aq Na<sub>2</sub> S<sub>2</sub>O<sub>3</sub> and water, dried

 $(Na_2SO_4)$  and concentrated under reduced pressure to give crude product, which was purified over SiO<sub>2</sub> using hexane-EtOAc (5:1) as eluant to furnish pure 18 (1.1 g, 74%); colorless solid; m.p. 78–80°C;  $[\alpha]_D^{25}$  –35.6 (c 1.5, CHCl<sub>3</sub>); IR (KBr): 2926, 2363, 1752, 1707, 1652, 1510, 1458, 1374, 1223, 1090, 698 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): § 7.39-7.15 (m, 40 H, Ar-H), 7.06-7.02 (m, 2 H, Ar-H), 6.90 (d, J=9.0 Hz, 2 H, Ar-H), 6.78–6.70 (m, 4 H, Ar-H), 5.51 (s, 1 H, PhCH), 5.30 (d, J=1.7 Hz, 1 H, H-1<sub>A</sub>), 5.28 (s, 1 H, PhCH), 5.08 (br s, 1 H, H-1<sub>D</sub>), 5.05 (br s, 1 H, H-1<sub>F</sub>), 5.04–5.0 (m, 2 H, H-2<sub>E</sub> and PhCH<sub>2</sub>), 4.97 (d, J=7.8 Hz, 1 H, H-1<sub>B</sub>), 4.94–4.91 (m, 1 H, H-3<sub>E</sub>), 4.90 (d, J=8.1 Hz, 1 H, H-1<sub>C</sub>), 4.89–4.76 (m, 6 H, PhCH<sub>2</sub>), 4.70 (d, J=7.2 Hz, 1 H, H-1<sub>E</sub>), 4.67–4.61 (m, 3 H, H-4<sub>E</sub> and PhCH<sub>2</sub>), 4.60–4.43 (m, 4 H, PhCH<sub>2</sub>), 4.38–4.20 (m, 6 H, H-3<sub>A</sub>, H-3<sub>B</sub>, H-3<sub>D</sub>, H-3<sub>F</sub>, H-6<sub>aC</sub>, PhCH<sub>2</sub>), 4.06–3.92 (m, 3 H,  $H-2_A$ ,  $H-3_C$ ,  $H-4_F$ ), 3.88–3.72 (m, 3 H,  $H-2_D$ ,  $H-5_A$  and H-6<sub>bC</sub>), 3.74 (s, 3 H, OCH<sub>3</sub>), 3.69 (s, 3 H, OCH<sub>3</sub>), 3.67–3.57 (m, 6 H, H-2<sub>F</sub>, H-4<sub>B</sub>, H-5<sub>D</sub>, H-5<sub>aE</sub>, H-5<sub>F</sub> and H-6<sub>aB</sub>), 3.55-3.38 (m, 5 H, H-2<sub>B</sub>, H-2<sub>C</sub>, H-4<sub>A</sub>, H-4<sub>D</sub> and H-5<sub>bE</sub>), 3.35 (s, 3 H, OCH<sub>3</sub>), 3.34–3.23 (m, 2 H, H-4<sub>C</sub> and H-5<sub>B</sub>), 3.22–3.14 (m, 1 H, H-5<sub>C</sub>), 2.96–2.88 (m, 1 H, H-6<sub>bB</sub>), 2.04, 2.01, 2.0 (3 s, 9 H, 3 COCH<sub>3</sub>), 1.30 (d, J=6.1 Hz, 3 H, CCH<sub>3</sub>), 1.25 (d, J=6.1 Hz, 3 H, CCH<sub>3</sub>), 0.81 (d, J=6.0 Hz, 3 H, CCH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 169.4, 169.1, 168.6 (3 COCH<sub>3</sub>), 159.6–113.4 (Ar-C), 103.3 (C-1<sub>C</sub>), 102.0 (C-1<sub>B</sub>), 101.2 (2 C, 2 PhCH), 100.7 (C-1<sub>E</sub>), 99.4 (C-1<sub>E</sub>), 98.7  $(C-1_D)$ , 97.1  $(C-1_A)$ , 83.2  $(C-2_F)$ , 81.8  $(C-5_F)$ , 81.6  $(C-2_C)$ , 81.3 (2 C, C-4<sub>A</sub> and C-4<sub>D</sub>), 80.9 (C-2<sub>B</sub>), 80.2 (C-5<sub>C</sub>), 79.3 (C-4<sub>B</sub>), 78.9 (C-2<sub>D</sub>), 78.4 (C-2<sub>A</sub>), 77.4 (C-5<sub>B</sub>), 77.2 (C-3<sub>B</sub>), 76.7 (2 C, C-3<sub>A</sub> and C-3<sub>C</sub>), 76.1 (C-5<sub>D</sub>), 74.8 (3 C, PhCH<sub>2</sub>), 74.3 (C-3<sub>F</sub>), 74.2 (PhCH<sub>2</sub>), 73.8 (PhCH<sub>2</sub>), 73.5 (PhCH<sub>2</sub>), 71.9 (PhCH<sub>2</sub>), 70.9 (C-3<sub>D</sub>), 70.7 (C-4<sub>E</sub>), 68.6 (2 C, C-5<sub>E</sub>) and C-6<sub>C</sub>), 68.5 (2 C, C-2<sub>E</sub> and C-5<sub>A</sub>), 68.4 (C-3<sub>E</sub>), 66.1 (C-4<sub>F</sub>), 65.8 (C-4<sub>C</sub>), 61.2 (C-6<sub>B</sub>), 57.3 (OCH<sub>3</sub>), 55.3, 54.8 (2 OCH<sub>3</sub>); ESI-MS: m/z=2012.8 [M+NH<sub>4</sub>]<sup>+</sup>; Anal. Calcd. for C113H126O32 (1994.82): C, 67.99; H, 6.36; found: C, 67.78; H, 6.60.

4-Methoxyphenyl (4-O-benzyl-3-O-methyl- $\alpha$ -L-rhamnopyranosyl)-(1 $\rightarrow$ 3)-[(2,3,4-tri-O-acetyl- $\beta$ -D-xylopyranosyl)-(1 $\rightarrow$ 2)]-(4-O-benzyl- $\alpha$ -L-rhamnopyranosyl)-(1 $\rightarrow$ 3)-(2-Obenzyl-4,6-O-benzylidene- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-(2-O-benzyl-4,6-O-benzylidine- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 3)-2,4-di-O-benzyl- $\alpha$ -L-rhamnopyranoside (19) To a solution of compound 18 (1 g, 0.5 mmol) in CH<sub>2</sub>Cl<sub>2</sub> and water (20 mL, 1:1), was added DDQ (140 mg, 0.6 mmol) and the reaction mixture was stirred at room temperature for 2 h and diluted with CH<sub>2</sub>Cl<sub>2</sub> (30 mL). The organic layer was washed successively with satd. aq NaHCO<sub>3</sub> and water, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure to give the crude product, which was purified over SiO<sub>2</sub> using hexane-EtOAc (5:1) as eluant to furnish pure 19 (750 mg, 80%); colorless solid; m.p. 98–100°C;  $[\alpha]_D^{25}$  –51.6 (c 1.5, CHCl<sub>3</sub>); IR (KBr): 3021, 2363, 1757, 1593, 1372, 1216, 1087, 760, 670 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.49– 7.06 (m, 40 H, Ar-H), 6.90 (d, J=9.0 Hz, 2 H, Ar-H), 6.77 (d, J=9.0 Hz, 2 H, Ar-H), 5.50 (s, 1 H, PhCH), 5.29 (br s, 1 H, H-1<sub>A</sub>), 5.24 (s, 1 H, PhCH), 5.08 (br s, 2 H, H-1<sub>D</sub> and H-1<sub>F</sub>), 5.04–4.99 (m, 3 H, H-1<sub>B</sub>, H-1<sub>C</sub> and PhCH<sub>2</sub>), 4.98– 4.94 (m, 1 H, H-2<sub>E</sub>), 4.90–4.80 (m, 5 H, PhCH<sub>2</sub>), 4.77–4.60 (m, 4 H, H-1<sub>E</sub> and PhC $H_2$ ), 4.58–4.45 (m, 4 H, H-3<sub>E</sub>, H-4<sub>E</sub> and PhCH<sub>2</sub>), 4.38–4.30 (m, 4 H, H-3<sub>A</sub>, H-3<sub>D</sub>, H-3<sub>F</sub> and PhCH<sub>2</sub>), 4.28–4.19 (m, 1 H, H-6<sub>aC</sub>), 4.05–3.90 (m, 5 H, H-2<sub>A</sub>, H-2<sub>F</sub>, H-3<sub>B</sub>, H-3<sub>C</sub> and H-4<sub>B</sub>), 3.88-3.76 (m, 4 H, H-2<sub>D</sub>, H-4<sub>F</sub>, H-5<sub>A</sub>, H-6<sub>bC</sub>), 3.74 (s, 3 H, OCH<sub>3</sub>), 3.72–3.52 (m, 4 H, H-5<sub>D</sub>, H-5<sub>F</sub>, H-5<sub>aE</sub> and H-6<sub>aB</sub>), 3.50–3.40 (m, 4 H, H-2<sub>B</sub>, H-2<sub>C</sub>, H-4<sub>A</sub> and H-5<sub>bE</sub>), 3.39 (s, 3 H, OCH<sub>3</sub>), 3.38-3.26 (m, 3 H, H-4<sub>C</sub>, H-4<sub>D</sub> and H-5<sub>B</sub>), 3.25-3.16 (m, 1 H, H-5<sub>C</sub>), 2.97–2.86 (m, 1 H, H-6<sub>bB</sub>), 2.01, 1.96 (2 s, 9 H, 3 COCH<sub>3</sub>), 1.30 (d, J=6.0 Hz, 3 H, CCH<sub>3</sub>), 1.25 (d, J= 6.0 Hz, 3 H, CCH<sub>3</sub>), 0.85 (d, J=6.0 Hz, 1 H, CCH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 169.2, 169.0, 168.6 (3 COCH<sub>3</sub>), 154.8-117.5 (Ar-C), 103.2 (C-1<sub>C</sub>), 102.0 (C-1<sub>B</sub>), 101.3 (PhCH), 101.2 (PhCH), 100.7 (2 C, C-1<sub>E</sub> and C-1<sub>F</sub>), 98.7 (C-1<sub>D</sub>), 97.1 (C-1<sub>A</sub>), 83.5 (C-5<sub>C</sub>), 83.2 (C-2<sub>F</sub>), 81.9 (C-2<sub>C</sub>), 81.5 (C-4<sub>A</sub>), 81.4 (C-4<sub>D</sub>), 81.0 (C-2<sub>B</sub>), 79.6 (C-5<sub>F</sub>), 79.3 (C-4<sub>B</sub>), 79.0 (C-2<sub>D</sub>), 78.4 (C-2<sub>A</sub>), 77.5 (2 C, C-3<sub>B</sub> and C-5<sub>B</sub>), 76.9 (C-3<sub>C</sub>), 76.2 (C-3<sub>A</sub>), 74.8 (2 C, PhCH<sub>2</sub>), 74.7 (PhCH<sub>2</sub>), 74.3 (PhCH<sub>2</sub>), 73.8 (PhCH<sub>2</sub>), 73.5 (PhCH<sub>2</sub>), 70.7 (C-5<sub>D</sub>), 70.6 (C-3<sub>F</sub>), 68.7 (2 C, C-5<sub>E</sub> and C-6<sub>C</sub>), 68.5 (2 C, C-3<sub>D</sub> and C-4<sub>E</sub>), 68.0 (C-2<sub>E</sub>), 67.7 (C-3<sub>E</sub>), 67.6 (C-5<sub>A</sub>), 66.0 (C-4<sub>F</sub>), 65.8 (C-4<sub>C</sub>), 61.2 (C-6<sub>B</sub>), 57.1 (OCH<sub>3</sub>), 55.3 (OCH<sub>3</sub>), 20.7, 20.5 (2 C) (3 COCH<sub>3</sub>), 18.2, 17.8, 17.2 (3 CCH<sub>3</sub>); ESI-MS: m/z=1897.8 [M+Na]<sup>+</sup>; Anal. Calcd. for C<sub>105</sub>H<sub>118</sub>O<sub>31</sub> (1874.76): C, 67.22; H, 6.34; found: C, 67.0; H, 6.58.

4-Methoxyphenyl (2,3,4-tri-O-acetyl- $\alpha$ -L-rhamnopyranosyl)- $(1 \rightarrow 2)$ -(4-O-benzyl-3-O-methyl- $\alpha$ -L-rhamnopyranosyl)- $(1 \rightarrow 3)$ - $[(2, 3, 4-tri-O-acetyl-\beta-D-xylopyranosyl)$ - $(1\rightarrow 2)$ ]- $(4-O-benzyl-\alpha-L-rhamnopyranosyl)-(1\rightarrow 3)-(2-O$  $benzyl-4, 6-O-benzylidene-\beta-D-glucopyranosyl)-(1\rightarrow 3) (2-O-benzyl-4, 6-O-benzylidine-\beta-D-glucopyranosyl)$ - $(1 \rightarrow 3)$ -2,4-di-O-benzyl- $\alpha$ -L-rhamnopyranoside (20) To a solution of compound 19 (700 mg, 0.37 mmol) and compound 4 (190 mg, 0.57 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (20 mL) was added MS 4 Å (3 g) and the reaction mixture was stirred at room temperature for 30 min under argon. The reaction mixture was cooled to  $-30^{\circ}$ C and Niodosuccinimide (150 mg, 0.66 mmol) and TMSOTf (5  $\mu$ L, 0.03 mmol) were added to it. The reaction mixture was stirred at the same temperature for 30 min and quenched with Et<sub>3</sub>N (50 µL). The reaction mixture was filtered and washed with CH<sub>2</sub>Cl<sub>2</sub> (30 mL). The organic layer was successively washed with 10% aq Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> and water,

dried  $(Na_2SO_4)$  and concentrated under reduced pressure to give crude product, which was purified over SiO<sub>2</sub> using hexane-EtOAc (5:1) as eluant to furnish pure 20 (675 mg, 85%); colorless solid; m.p. 95–97°C;  $[\alpha]_D^{25}$  –45 (c 1.5, CHCl<sub>3</sub>); IR (KBr): 3021, 2925, 2365, 1753, 1662, 1600, 1369, 1219, 1085, 761 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>): δ 7.48-7.02 (m, 40 H, Ar-H), 6.89 (d, J=9.0 Hz, 2 H, Ar-H), 6.78 (d, J=9.0 Hz, 2 H, Ar-H), 5.30 (s, 1 h, PhCH), 5.40-5.20 (m, 4 H, H-1<sub>A</sub>, H-2<sub>G</sub>, H-3<sub>G</sub> and PhCH), 5.14-4.78 (m, 10 H, H-1<sub>B</sub>, H-1<sub>C</sub>, H-1<sub>D</sub>, H-1<sub>F</sub>, H-1<sub>G</sub>, H-2<sub>F</sub>, H-4<sub>G</sub> and PhCH<sub>2</sub>), 4.75–4.60 (m, 7 H, H-1<sub>E</sub>, H-3<sub>E</sub>, H-4<sub>E</sub> and PhCH<sub>2</sub>), 4.58–4.43 (m, 4 H, PhCH<sub>2</sub>), 4.40–4.20 (m, 5 H, H-3<sub>A</sub>, H-3<sub>D</sub>, H-3<sub>F</sub>, H-6<sub>aC</sub> and PhCH<sub>2</sub>), 4.13–4.07 (m, 1 H, H-3<sub>B</sub>), 4.05–3.90 (m, 5 H, H-2<sub>A</sub>, H-2<sub>D</sub>, H-2<sub>F</sub>, H-3<sub>C</sub> and H-4<sub>B</sub>), 3.88–3.70 (m, 4 H, H-4<sub>F</sub>, H-5<sub>A</sub>, H-5<sub>G</sub> and H-6<sub>bC</sub>), 3.75 (s, 3 H, (OCH<sub>3</sub>), 3.68–3.50 (m, 6 H, H-4<sub>A</sub>, H-4<sub>D</sub>, H-5<sub>D</sub>, H-5<sub>F</sub>, H-5<sub>aE</sub> and H-6<sub>aB</sub>), 3.46 (s, 3 H, OCH<sub>3</sub>), 3.44-3.10 (m, 6 H, H-2<sub>B</sub>, H-2<sub>C</sub>, H-4<sub>C</sub>, H-5<sub>B</sub>, H-5<sub>C</sub> and H-5<sub>bE</sub>), 2.98-2.88 (m, 1 H, H-6<sub>bB</sub>), 2.05, 2.04, 2.02, 2.01, 2.00, 1.98 (6 s, 18 H, 6 COCH<sub>3</sub>), 1.31–1.20 (m, 9 H, 3 CCH<sub>3</sub>), 0.87 (d, J=6.0 Hz, CCH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 169.9-168.4 (6 COCH<sub>3</sub>), 154.8-114.5 (Ar-C), 102.2 (C-1<sub>C</sub>), 101.1 (C-1<sub>B</sub>), 100.2 (2 C, PhCH), 99.6 (C-1<sub>F</sub>), 99.4 (C-1<sub>E</sub>), 97.7 (2 C, C-1<sub>D</sub> and C-1<sub>G</sub>), 96.1 (C-1<sub>A</sub>), 83.2 (C-5<sub>C</sub>), 82.2 (C-2<sub>F</sub>), 81.4 (3 C, C-2<sub>C</sub>, C-4<sub>A</sub> and C-4<sub>D</sub>), 81.0 (C-5<sub>G</sub>), 80.0 (C-2<sub>B</sub>), 79.4 (C-5<sub>F</sub>), 79.1 (C-4<sub>B</sub>), 78.5 (C-2<sub>D</sub>), 77.7  $(C-2_A)$ , 77.6  $(C-3_B)$ , 77.2  $(C-5_B)$ , 76.8  $(C-3_C)$ , 76.4  $(C-3_A)$ , 75.2 (C-3<sub>F</sub>), 75.1 (PhCH<sub>2</sub>), 75.9 (PhCH<sub>2</sub>), 74.7 (PhCH<sub>2</sub>), 74.4 (PhCH<sub>2</sub>), 74.0 (PhCH<sub>2</sub>), 73.6 (PhCH<sub>2</sub>), 70.9 (3 C, C-3<sub>D</sub>, C-4<sub>E</sub> and C-5<sub>D</sub>), 70.8 (C-4<sub>G</sub>), 69.7 (C-2<sub>G</sub>), 68.9 (C-3<sub>G</sub>), 68.8 (2 C, C-5<sub>E</sub> and C-6<sub>C</sub>), 68.5 (2 C, C-2<sub>E</sub> and C-3<sub>E</sub>), 67.8 (C-5<sub>A</sub>), 66.8 (C-4<sub>F</sub>), 66.2 (C-4<sub>C</sub>), 61.5 (C-6<sub>B</sub>), 57.9 (OCH<sub>3</sub>), 55.4 (OCH<sub>3</sub>), 20.8–20.6 (6 COCH<sub>3</sub>), 18.3, 17.9, 17.2, 17.1 (4 CCH<sub>3</sub>); ESI-MS: m/z=2165.0 [M+NH<sub>4</sub>]<sup>+</sup>; Anal. Calcd. for C<sub>117</sub>H<sub>134</sub>O<sub>38</sub> (2146.85): C, 65.41; H, 6.29; found: C, 65.22; H, 6.55.

4-Methoxyphenyl ( $\alpha$ -L-rhamnopyranosyl)-(1 $\rightarrow$ 2)-(3-O-meth $vl-\alpha-L-rhamnopyranosyl)-(1\rightarrow 3)-[(\beta-D-xylopyranosyl) (1\rightarrow 2)]-(\alpha-L-rhamnopyranosyl)-(1\rightarrow 3)-(\beta-D-glucopyranosyl) (1 \rightarrow 3)$ - $(\beta$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ - $\alpha$ -L-rhamnopyranoside (1) A solution of compound 20 (650 mg, 0.3 mmol) in 0.1 M CH<sub>3</sub>ONa in CH<sub>3</sub>OH (10 mL) was allowed to stir at room temperature for 2 h. The reaction mixture was neutralized with Dowex 50 W X-8 (H<sup>+</sup>) resin, filtered and concentrated. To a solution of the crude product in CH<sub>3</sub>OH (10 mL) was added 20% Pd(OH)<sub>2</sub>/C (150 mg) and the reaction mixture was allowed to stir at room temperature under a positive pressure of hydrogen for 24 h. The reaction mixture was filtered through a Celite® bed and evaporated to dryness to give heptasaccharide 1, which was purified through a Sephadex LH-20 column using CH<sub>3</sub>OH-water (8:1) as eluant (270 mg, 76%); white powder;  $[\alpha]_{D}^{25}$  -52.3

(c 1.0, H<sub>2</sub>O); IR (KBr); 2926, 2373, 1759, 1658, 1550, 1467, 1429, 1370, 1045, 679 cm<sup>-1</sup>; <sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O): δ 7.0 (d, J=9.0 Hz, 2 H, Ar-H), 6.88 (d, J=9.0 Hz, 2 H, Ar-H), 5.39 (br s, 1 H, H-1<sub>G</sub>), 5.33 (br s, 1 H, H-1<sub>A</sub>), 5.26 (br s, 1 H, H-1<sub>D</sub>), 4.92 (br s, 1 H, H-1<sub>F</sub>), 4.70 (d, J=7.5 Hz, 1 H, H-1<sub>B</sub>), 4.63 (d, J=7.5 Hz, 1 H, H-1<sub>C</sub>), 4.54  $(d, J=7.2 \text{ Hz}, 1 \text{ H}, \text{H}-1_{\text{E}}), 4.30 \text{ (br s}, 1 \text{ H}, \text{H}-2_{\text{A}}), 4.23 \text{ (br s}, 1 \text{ H}, \text{H}-2_{\text{A}})$ 1 H, H-2<sub>D</sub>), 4.07–4.03 (m, 2 H, H-2<sub>G</sub> and H-3<sub>A</sub>), 4.0–3.82 (m, 6 H, H-2<sub>F</sub>, H-4<sub>B</sub>, H-4<sub>G</sub>, H-5<sub>aE</sub> and H-6<sub>a,bB</sub>), 3.80–3.78 (m, 1 H, H-3<sub>F</sub>), 3.76 (s, 3 H, OCH<sub>3</sub>), 3.70–3.61 (m, 5 H,  $H-3_E$ ,  $H-3_G$ ,  $H-4_F$  and  $H-6_{a,bC}$ ), 3.60–3.51 (m, 7 H,  $H-2_B$ , H-3<sub>B</sub>, H-3<sub>C</sub>, H-3<sub>D</sub>, H-4<sub>A</sub>, H-4<sub>D</sub>, H-5<sub>G</sub>), 3.50 (s, 3 H, OCH<sub>3</sub>), 3.45–3.30 (m, 10 H, H-2<sub>C</sub>, H-2<sub>E</sub>, H-4<sub>C</sub>, H-4<sub>E</sub>, H-5<sub>A</sub>, H-5<sub>B</sub>, H-5<sub>C</sub>, H-5<sub>D</sub>, H-5<sub>bE</sub>, H-5<sub>F</sub>), 1.36–1.27 (m, 9 H, 3 CCH<sub>3</sub>), 0.91 (d, J=6.0 Hz, 3 H, CCH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O): δ 155.0–114.2 (Ar-C), 105.0 (C-1<sub>E</sub>), 103.9 (C-1<sub>B</sub>), 103.5 (C-1<sub>C</sub>), 101.7 (C-1<sub>F</sub>), 100.7 (C-1<sub>D</sub>), 100.0 (C-1<sub>G</sub>), 99.0  $(C-1_A)$ , 85.9  $(C-3_G)$ , 82.1  $(C-3_D)$ , 81.2  $(C-2_G)$ , 80.4  $(C-5_G)$ , 78.8 (C-3<sub>A</sub>), 76.5 (C-4<sub>C</sub>), 76.0 (2 C, C-2<sub>C</sub>, C-4<sub>G</sub>), 75.9  $(C-4_E)$ , 74.7  $(C-5_F)$ , 73.6  $(C-4_A)$ , 73.3  $(C-2_D)$ , 73.1  $(C-3_C)$ , 72.4 (2 C, C-2<sub>B</sub>, C-4<sub>B</sub>), 71.8 (C-5<sub>B</sub>), 71.2 (C-2<sub>A</sub>), 70.8  $(C-2_E)$ , 70.7  $(C-5_A)$ , 70.0  $(C-4_E)$ , 69.7  $(C-3_B)$ , 69.2  $(C-3_E)$ , 68.8 (2 C, C-5<sub>C</sub>, C-5<sub>D</sub>), 68.7 (C-3<sub>F</sub>), 68.5 (C-4<sub>D</sub>), 68.1 (C-2<sub>F</sub>), 65.4 (C-5<sub>E</sub>), 61.1 (C-6<sub>B</sub>), 60.6 (C-6<sub>C</sub>), 56.7 (OCH<sub>3</sub>), 54.6 (OCH<sub>3</sub>), 16.8, 16.6, 16.4, 16.3 (4 CCH<sub>3</sub>); ESI-MS: m/z=1201.5 [M+Na]<sup>+</sup>; Anal. Calcd. for C<sub>49</sub>H<sub>78</sub>O<sub>32</sub> (1178.45): C, 49.91; H, 6.67; found: C, 49.67; H, 6.96.

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